

SEQUENCE ANALYSIS SUGGESTS POSITIVE SELECTION ON THE BOVINE PRODYNORPHIN GENE



ANÁLISIS DE SECUENCIAS GENÓMICAS SUGIEREN QUE EL GEN DE LA PRODINORFINA ESTÁ BAJO SELECCIÓN POSITIVA EN BOVINOS

Suqueli García M.F.¹, Castellote M.A.², Corva P.M.¹

ABSTRACT

Dynorphin A is an endogenous opioid peptide that is part of the KNDy system in the hypothalamus of mammals. This peptide acts as an inhibitor of the GnRH pulse generation, thus regulating the onset of puberty and reproductive cycles. The *PDYN* gene encodes the propeptide Prodynorphin, the precursor of Dynorphin A. Despite its physiological relevance, *PDYN* has not emerged as a candidate gene associated with puberty in genomic association studies conducted in cattle. The present work aimed to search for signatures of selection on the *PDYN* gene among cattle breeds. To this, the whole genome sequences from 57 samples of ten cattle breeds were used. The samples were grouped based on breed selection history and their productive differences, particularly in terms of sexual precocity. The population structure was analyzed using Principal Component Analyses. To evidence recent selection processes, neutrality tests, such as Tajima's D and Fu & Li's F* and D* were performed in defined functional regions of *PDYN*. The putative promoter of *PDYN* showed a population structure that is in agreement with the criteria considered to make the groups. In that region, neutrality tests were consistently negative and resulted in statistically significant for the dairy breeds. Also, these breeds exhibited less variability in the haplotype analyses than the others. The results presented here suggest that regulatory regions of *PDYN* could be under positive selection, particularly in dairy breeds.

Key words: reproduction; KNDy neurons; Dynorphin; signatures of selection.

RESUMEN

Dinorfina A es un péptido opioide endógeno que forma parte del sistema KNDy en el hipotálamo de mamíferos. Este péptido actúa como inhibidor de la generación de los pulsos de GnRH, regulando así el inicio de la pubertad y los ciclos reproductivos. El gen *PDYN* codifica el propéptido Prodinorfina, precursor de Dinorfina A. A pesar de su relevancia fisiológica, *PDYN* no ha surgido como gen candidato asociado a pubertad en estudios de asociación genómicos en bovinos. El presente trabajo tuvo como objetivo buscar huellas de selección en el gen *PDYN* entre diferentes razas bovinas. Para alcanzarlo se utilizaron secuencias genómicas de 57 muestras de diez razas bovinas. Las muestras fueron agrupadas considerando la historia de selección y las diferencias productivas entre razas, particularmente en términos de precocidad sexual. La estructura poblacional fue analizada usando análisis de componentes principales. Para evidenciar procesos de selección recientes se realizaron pruebas de neutralidad, tales como D de Tajima y F* y D* de Fu & Li, en diferentes regiones funcionales de *PDYN*. El promotor putativo de *PDYN* mostró una estructura poblacional que es consistente con los criterios usados para agrupar las razas. En esa región, las pruebas de neutralidad fueron consistentemente negativas y estadísticamente significativas en las razas lecheras. Además, estas razas también exhibieron menor variabilidad en los análisis de haplotipos que las demás razas. Los resultados presentados aquí sugieren que regiones regulatorias de *PDYN* estarían bajo selección positiva, particularmente en razas bovinas lecheras.

Palabras clave: reproducción; neuronas KNDy; Dinorfina; huellas de selección.

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INTRODUCTION

Reproductive efficiency is one of the most valued aspects of animal production. In the particular case of beef and dairy cattle, reproduction-associated traits, especially of females, are included in the selection objective of the most modern breeds and they have received much attention in genetic and genomic studies.

Starting with puberty and then cyclically repeated along productive life, the reproductive process in females requires the release of the gonadotropin-releasing hormone (GnRH) from the hypothalamus, which in turn is needed for the secretion of gonadotropins from the anterior pituitary gland (Amstalden and Williams, 2015; Atkins *et al.*, 2013). Research devoted to understanding the mechanisms underlying the onset of puberty and the regulation of subsequent reproductive cycles in different mammal species contributed to building the “KNDy hypothesis” (Smith *et al.*, 2014). According to this hypothesis three peptides, Kisspeptin, Neurokinin B, and Dynorphin A, are co-expressed in the same group of neurons in specific regions of the hypothalamus (Goodman *et al.*, 2007). These peptides would have the role of a “pulse generator” that drives GnRH secretion. Neurokinin B and Dynorphin A have stimulatory and inhibitory effects on Kisspeptin expression, respectively. Moreover, these neurons also express the Neurokinin B receptor and the Dynorphin A receptor, but not the Kisspeptin receptor that is expressed in GnRH neurons (Weems *et al.*, 2018). The regulatory system involving the KNDy neurons would be sensitive to sex steroid feedback and different regulatory cues (*e.g.* energy balance, lactation, photoperiod, stress) but the specific mechanisms involved are not clear (Lehman *et al.*, 2010).

The role of Dynorphin A in the KNDy neurons is consistent with previous evidence showing an inhibitory effect of endogenous opioid peptides on reproduction (Malven, 1986). Suckling increases levels of endogenous opioid peptides in the brain, making the hypothalamus more sensitive to the negative feedback from estrogen and decreasing the production of GnRH (Squires, 2010).

Despite the role of Kisspeptin, Neurokinin B and Dynorphin A in the regulation of the reproductive cycle, the corresponding genes (*KISS1*, *TAC3*, and *PDYN*) and their receptors (*KISS1R*, *TACR3*, and *OPRK1*) have not been analyzed as functional or positional candidates in cattle. The only known exception is the Dynorphin receptor (*OPRK1*), proposed as a candidate gene associated with sexual precocity in a GWAS for sexual precocity in Nellore cattle from Brazil (Irano *et al.*, 2016).

From the standpoint of animal breeding, the dissection of genetic variation at the level of the KNDy system could be of help to understand differences in reproductive performance among cattle breeds, and in turn, to improve management strategies. As an example, we have evaluated the reproductive efficiency of Angus

and Argentinean Creole females (Corva *et al.*, 1995). The Creole is a local beef breed that has been much less selected than Angus, but it is appreciated due to its rusticity and some other favorable traits such as calving ease. The superiority of Angus over Creole in reproductive performance was noticed starting with age at puberty (Pardo *et al.*, 2018) and other differences between the two breeds were detected in subsequent reproductive cycles (Corva *et al.*, 1995). While body weight change during the breeding season was a key factor defining pregnancy rate in Angus females, lactation and the interaction with the offspring was the most limiting factor in Creole females. The marked difference between European (*Bos taurus*) and Indicine (*Bos indicus*) breeds in sexual precocity is another well-known example of genetic variation in reproductive performance (Freetly *et al.*, 2011; Laster *et al.*, 1979) even when were evaluated in similar restrictive environmental conditions (Meirelles *et al.*, 1994; Ferraz Jr. *et al.*, 2018).

Taking together the evidences about the inhibitory effect of endogenous opioid peptides on reproduction and the recently proposed role of Dynorphin A in KNDy neurons, we hypothesized that *PDYN*, the gene that codes for the propeptide Prodynorphin, which is cleaved to produce several opioid peptides including Dynorphin A (Day *et al.*, 1998, Figure 1), could be under selection in cattle. This selection probably resulting in the relaxation of the effects of different cues that convey to the hypothalamus to regulate the onset of reproductive activity. To test this hypothesis, we compared the sequence of the *PDYN* gene from different cattle breeds, taking advantage of the availability of whole genome sequences.

MATERIALS AND METHODS

Retrieval and processing of *PDYN* genomic sequences

The genomic sequence of the Bovine Reference Genome (ARS-UCD1.2) corresponding to a region of BTA13 spanning the *PDYN* gene (GenBank accession: NC_037340.1: 53,130,235 - 53,148,819 bp) was downloaded from the NCBI site (<http://www.ncbi.nlm.nih.gov>). This DNA sequence was used as a reference to retrieve the corresponding genomic sequence of *Bos indicus* (GenBank accession: NC_032662.1: 53,688,047 - 53,707,634 bp) and also the genomic sequences of a panel of individuals from selected bovine breeds as it is described below. The sequenced genomes of selected animals had already been aligned to the Bovine Reference Genome UMD 3.1.1 (Merchant *et al.*, 2014; Zimin *et al.*, 2009). Sequence alignment confirmed that apart from a change in relative coordinates, the region of interest resulted in identical between UMD 3.1.1 and ARS-UCD1.2, the latest version of the Bovine Reference Genome (www.bovinegenome.org).

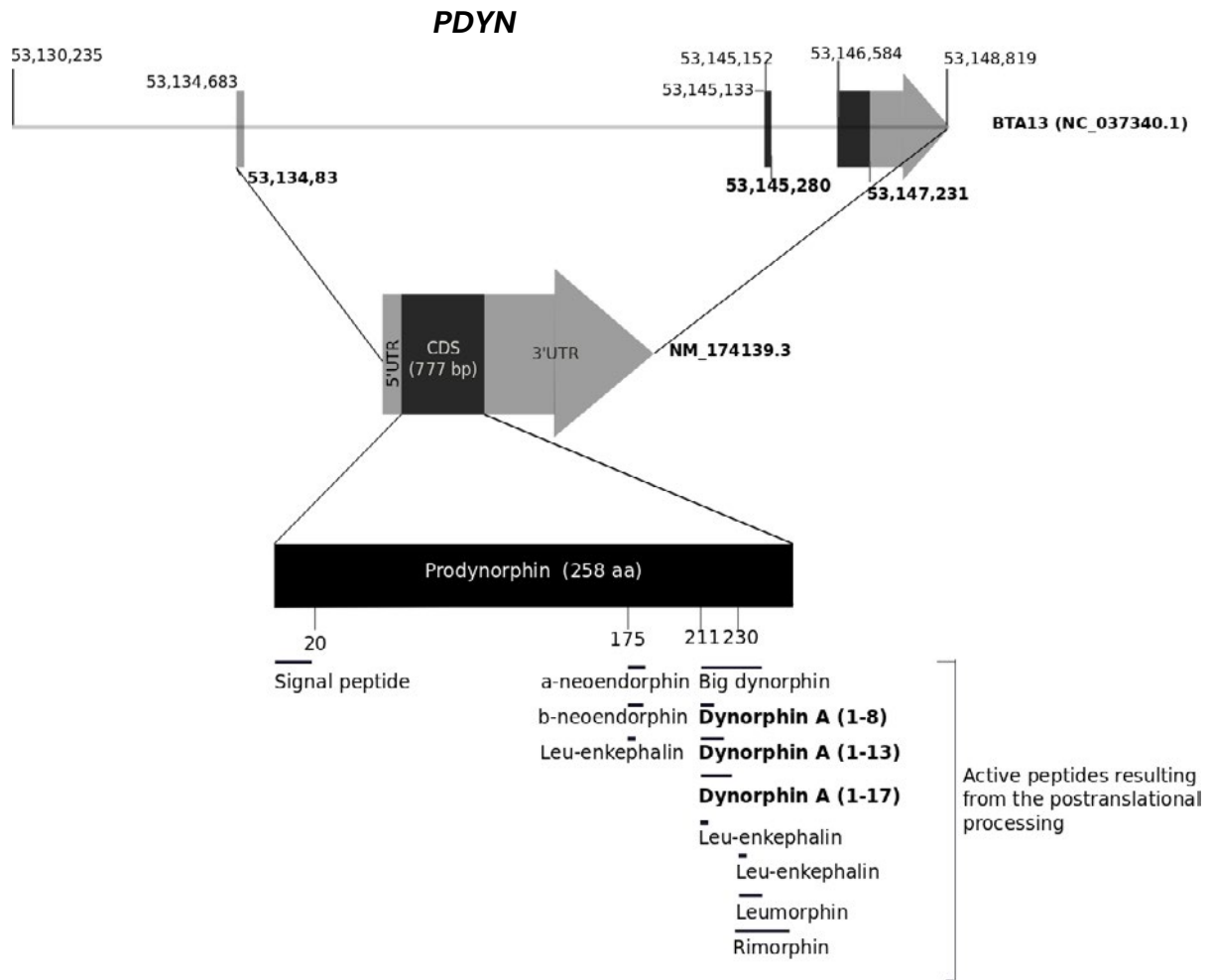


Figure 1. Structure of the bovine *PDYN* gene (Top), the corresponding transcript (Genbank sequence: NM_174139.3) (Middle) and the translated propeptide (Bottom). The different functional peptides resulting from proteolytic cleavage by the enzyme Prohormone convertase 2 are also indicated in the figure.

The whole panel of genomic sequences used for the analyses included: The reference *Bos taurus* and *Bos indicus* genomes, corresponding to the Hereford and Nellore breeds respectively; one individual representing extinct wild cattle (*Auroch*, *Bos primigenius*, Park *et al.*, 2015) and individuals from the “1000 Bull Genomes Project” (<http://www.1000bullgenomes.com/>) or the U.S. Meat Animal Research Center (USMARC) Beef Cattle Diversity Panel 2.9 (MBCDPv2.9, Heaton *et al.*, 2016). Selected breeds were: Holstein (n=13), Jersey (n=11), Angus (n=10), Limousine (n=6), Corriente (n=4), Longhorn (n=3) and Brahman (n=7). In the first experiment, the average sequencing depth was 8.3X (Daetwyler *et al.*, 2014) while in the experiment of Heaton *et al.* (2016) it was 14.8X. Aligned genomic sequences (in bam format) were downloaded from the Sequence Read Archive (SRA) division of NCBI (<http://www.ncbi.nlm.nih.gov/sra>) and the USMARC website (<https://www.ars.usda.gov/plains-area/clay-center-ne/marc/wgs/bovref/>), respectively. A detailed list of these genomic sequences is presented in Table 1. Based on breed selection history and well

known productive differences among bovine biotypes (particularly in terms of sexual precocity; Diskin and Kenny, 2014; Sartori *et al.*, 2016) four contrasting groups were defined: highly selected *Bos taurus* dairy (Holstein, Jersey, n=24) and beef breeds (Angus, Limousine, Hereford, n=17); less selected -hardy- *Bos taurus* breeds (Corriente, Longhorn, Auroch, n=8) and zebu breeds (Brahman, Nellore, n=8). These groups are hereinafter identified as “Dairy”, “Beef”, “Less selected” and “Zebu”, respectively.

Each genomic sequence in bam format was searched for variant sites using BCFtools software v1.5 (Li, 2011) with the command: “bcftools mpileup -f sequence.fasta input.bam | bcftools call -m -Ob | tabix | bcftools query -f ‘%POS\t%QUAL\t%ALT[\t%IUPACGT]\n’ -o output.txt” where “sequence.fasta” was the sequence of the Bovine Reference Genome UMD 3.1.1 used as a reference in the original alignment. Variants with a Phred quality score below 20 (base call accuracy above 99%) were manually discarded and a consensus sequence was obtained for each sample in fasta format using UNIX

Table 1. Genomic sequences used to characterize the bovine Prodynorphin gene. Each retrieved sequence was aligned to the region of BTA13 spanning the *PDYN* gene (Bovine Reference Genome ARS-UCD1.2, GenBank accession: NC_037340.1: 53,129,235–53,148,819).

| ID | Group | Breed | Whole genome coverage | <i>PDYN</i> region coverage |
|-------------|-------|------------------------|-----------------------|-----------------------------|
| NC_037340.1 | Beef | Hereford ¹ | | |
| SRR1262614 | Beef | Angus ² | 10.0X | 6.7X |
| SRR1262615 | Beef | Angus ² | 10.0X | 7.5X |
| SRR1262616 | Beef | Angus ² | 9.5X | 6.6X |
| SRR1262617 | Beef | Angus ² | 9.8X | 6.9X |
| SRR1262618 | Beef | Angus ² | 10.0X | 7.3X |
| SRR1262619 | Beef | Angus ² | 10.0X | 7.0X |
| SRR1262620 | Beef | Angus ² | 11.0X | 7.5X |
| SRR1262621 | Beef | Angus ² | 6.7X | 4.7X |
| SRR1262622 | Beef | Angus ² | 8.0X | 5.4X |
| SRR1262623 | Beef | Angus ² | 8.5X | 6.5X |
| 19989863 | Beef | Limousine ³ | 10.7X | 12.8X |
| 19999833 | Beef | Limousine ³ | 11.9X | 12.9X |
| 19999834 | Beef | Limousine ³ | 10.6X | 10.9X |
| 19999835 | Beef | Limousine ³ | 11.5X | 10.5X |
| 19999837 | Beef | Limousine ³ | 11.2X | 8.7X |
| 19999838 | Beef | Limousine ³ | 10.4X | 8.8X |
| SRR1262783 | Dairy | Holstein ² | 5.7X | 3.6X |
| SRR1188706 | Dairy | Holstein ² | 16.0X | 12.8X |
| SRR1262533 | Dairy | Holstein ² | 6.6X | 3.4X |
| SRR1262536 | Dairy | Holstein ² | 6.4X | 4.5X |
| SRR1262538 | Dairy | Holstein ² | 6.3X | 4.6X |
| SRR1262539 | Dairy | Holstein ² | 6.1X | 3.2X |
| SRR1262660 | Dairy | Holstein ² | 5.6X | 4.1X |
| SRR1262661 | Dairy | Holstein ² | 13.0X | 12.5X |
| SRR1262662 | Dairy | Holstein ² | 5.7X | 5.24X |
| SRR1262663 | Dairy | Holstein ² | 10.0X | 8.1X |
| SRR1262664 | Dairy | Holstein ² | 14X | 10.7X |
| SRR1262782 | Dairy | Holstein ² | 12.0X | 9.3X |
| SRR1262785 | Dairy | Holstein ² | 15.0X | 12.4X |

ID: File identification in the original database. Dairy: Highly selected dairy breeds (*Bos taurus*); Beef: Highly selected beef breeds (*Bos taurus*); Less selected: less selected breeds (*Bos taurus*), includes the Auroch (*Bos primigenius*); Zebu: zebu breeds (*Bos indicus*). All samples were aligned to the genomic region of BTA13 spanning the *PDYN* gene and only this region of each sample was used for the analyses. ¹Bovine Reference Genome ARS-UCD1.2, GenBank accession: NC_037340.1: 53,130,235–53,148,819). ²"1000 Bull Genome Project" Run 2, (<http://www.1000bullgenomes.com/>). ³U.S. Meat Animal Research Center (USMARC) Beef Cattle Diversity Panel 2.9 (MBCDPv2.9, Heaton et al., 2016). ⁴*Bos primigenius* archeological sample (Park et al., 2015). ⁵*Bos indicus* Reference Genome Bos_indicus_1.0 (GenBank accession: NC_032662.1). *Average depth of coverage obtained from their respective original database. #Average depth of coverage calculated by using Mosdepth software v 0.2.9 (Pedersen and Quinlan, 2018).

Table 1. (cont.) Genomic sequences used to characterize the bovine Prodynorphin gene. Each retrieved sequence was aligned to the region of BTA13 spanning the *PDYN* gene (Bovine Reference Genome ARS-UCD1.2, GenBank accession: NC_037340.1: 53,129,235–53,148,819).

| ID | Group | Breed | Whole genome coverage* | <i>PDYN</i> region coverage# |
|-------------|---------------|------------------------|------------------------|------------------------------|
| SRR1262789 | Dairy | Jersey ² | 8.8X | 6.0X |
| SRR1262790 | Dairy | Jersey ² | 8.3X | 6.2X |
| SRR1262791 | Dairy | Jersey ² | 9.3X | 7.0X |
| SRR1262792 | Dairy | Jersey ² | 6.9X | 5.4X |
| SRR1262793 | Dairy | Jersey ² | 14.0X | 10.7X |
| SRR1262794 | Dairy | Jersey ² | 6.4X | 5.1X |
| SRR1262795 | Dairy | Jersey ² | 7.1X | 6.5X |
| SRR1262796 | Dairy | Jersey ² | 6.1X | 3.7X |
| SRR1262797 | Dairy | Jersey ² | 11.0X | 8.9X |
| SRR1262798 | Dairy | Jersey ² | 13.0X | 11.3X |
| SRR1262803 | Dairy | Jersey ² | 12.0X | 12.0X |
| SRR2465682 | Less selected | Auroch ⁴ | 8.8X | 6.9X |
| 19202900 | Less selected | Corriente ³ | 11.3X | 10.1X |
| 19202901 | Less selected | Corriente ³ | 10.1X | 10.1X |
| 19202902 | Less selected | Corriente ³ | 12.9X | 11.9X |
| 19202903 | Less selected | Corriente ³ | 10.5X | 11.2X |
| 19999870 | Less selected | Longhorn ³ | 10.5X | 10.1X |
| 19999871 | Less selected | Longhorn ³ | 10.2X | 11.1X |
| 19999872 | Less selected | Longhorn ³ | 10.0X | 9.2X |
| NC_032662.1 | Zebu | Nellore ⁵ | | |
| 19919842 | Zebu | Brahman ³ | 19.6X | 21.5X |
| 19999811 | Zebu | Brahman ³ | 19.6X | 21.5X |
| 19999812 | Zebu | Brahman ³ | 10.0X | 10.2X |
| 19999813 | Zebu | Brahman ³ | 12.6X | 9.3X |
| 19999819 | Zebu | Brahman ³ | 12.1X | 12.3X |
| SRR3659494 | Zebu | Brahman ³ | 6.0X | 4.2X |
| SRR3659495 | Zebu | Brahman ³ | 5.3X | 3.9X |

ID: File identification in the original database. Dairy: Highly selected dairy breeds (*Bos taurus*); Beef: Highly selected beef breeds (*Bos taurus*); Less selected: less selected breeds (*Bos taurus*), includes the Auroch (*Bos primigenius*); Zebu: zebu breeds (*Bos indicus*). All samples were aligned to the genomic region of BTA13 spanning the *PDYN* gene and only this region of each sample was used for the analyses. ¹Bovine Reference Genome ARS-UCD1.2, GenBank accession: NC_037340.1: 53,130,235–53,148,819). ²"1000 Bull Genome Project" Run 2, (<http://www.1000bullgenomes.com/>). ³U.S. Meat Animal Research Center (USMARC) Beef Cattle Diversity Panel 2.9 (MBCDPv2.9, Heaton et al., 2016). ⁴*Bos primigenius* archeological sample (Park et al., 2015). ⁵*Bos indicus* Reference Genome Bos_indicus_1.0 (GenBank accession: NC_032662.1). *Average depth of coverage obtained from their respective original database. #Average depth of coverage calculated by using Mosdepth software v 0.2.9 (Pedersen and Quinlan, 2018).

commands. The average depth of coverage of each sample for the whole genome sequence was retrieved from the original database. On the other hand, the average depth of coverage of each sample for the *PDYN* gene region was calculated using Mosdepth software v 0.2.9 (Pedersen and Quinlan, 2018) (Table 1).

Haplotype reconstruction

The Prodynorphin cDNA was cloned by Jiang *et al.* (1997); Genbank accession U58500.1. This sequence agrees with accession NM_174139.3 from the annotation of the last version of the Bovine Reference Genome (ARS-UCD1.2), and therefore it was used in the present work as a reference of gene organization. In the comparative sequence analyses, the whole sequence and four functional regions of the mRNA were considered: a 1,000 bp fragment spanning the putative proximal promoter; the 5' untranslated region (5'UTR, 174 bp); the coding sequence (CDS, 777 bp); and the 3' untranslated region (3'UTR, 1,588 bp) (Figure 1).

The sequences of the breed panel were aligned online with the MAFFT software version 7 (<https://mafft.cbrc.jp/alignment/server/>) and the resulting alignment was manually edited using the Aliview software version 1.23 (Larsson, 2014). Then, haplotype phases from sequences with heterozygous sites were reconstructed with DnaSP version 6.10.03 (Rozas *et al.*, 2017) using the option fastPHASE (Scheet and Stephens, 2006) with default parameters. Haplotypes were reconstructed for the whole sequence and then trimmed into the functional regions as defined above.

The phylogenetic relationships among the haplotypes identified in the putative proximal promoter (1,000 bp) and in the coding sequence (CDS) of the *PDYN* gene, were inferred through a median-joining network analysis (Bandelt *et al.*, 1999) using the PopART software version 1.7 (Leigh and Bryant, 2015).

Principal component analyses (PCA)

To evaluate population structure and the genetic relationships among the four breed groups ("Dairy", "Beef", "Less selected" and "Zebu"), principal component analyses (PCA) were computed on haplotype frequencies of both, the putative proximal promoter region (1,000 bp) and the cDNA (complementary DNA, 2,539 bp), of *PDYN* using the 'prcomp' function of Rstudio v.3.4.4 (R Development Core Team, 2018). PCA results were plotted using the R package 'ggplot2' (Wickham, 2016).

Signatures of selection

To detect departures from the expectations of the neutral theory of evolution (Kimura, 1968) the neutrality tests Tajima's D (Tajima, 1989) and Fu & Li's F^* and D^* (Fu and Li, 1993) were conducted. Tajima's D compares the

estimate of DNA sequence variation based on the average pairwise distance between all sequences in the sample (ϕ), to the estimate of DNA sequence variation based on the observed number of segregating sites and the number of chromosomes in a sample (θ). Under neutrality, the means θ and ϕ should be approximately equal to each other. Therefore, the expected value of Tajima's D for populations conforming to a standard neutral model is zero. Significant deviations from zero indicate a skew in the allele frequency distribution relative to neutral expectations. Positive values of Tajima's D arise from an excess of intermediate frequency alleles and can result from population bottlenecks, structure and/or balancing selection. Negative values of Tajima's D indicate an excess of low frequency alleles and can result from population expansions or positive selection (Tajima, 1989; Biswas and Akey, 2006). On the other hand, Fu & Li's test makes the distinction between old and recent mutations as determined by where they occur on the branches of genealogies. The D^* and F^* statistics compare an estimate of the population mutation rate based on the number of derived variants seen only once in a sample (referred to as singletons) with ϕ or θ , respectively. Fu & Li's F^* and D^* values are negative when there is an excess of recent mutations and will be taken as evidence against the neutrality of mutations (Fu and Li, 1993; Biswas and Akey, 2006). These tests were conducted in the four defined cattle groups for the whole *PDYN* gene and also for each functional region defined above, using DnaSP version 6.10.03 (Rozas *et al.*, 2017).

RESULTS

Principal component analyses

The PCA conducted on the haplotype frequencies of the cDNA and the putative promoter of *PDYN* are presented in Figure 2. It can be seen that the distribution of the breed groups does not agree between both gene regions. In the case of the promoter, PC1 alone explains 79.7% of the variability. Breeds are distributed along this axis, in agreement with the criteria that were originally considered to make the groups (selection history and, particularly, sexual precocity) suggesting a population structure defined not only by breed isolation and genetic drift but probably also by selection. On the contrary, when haplotype frequencies of the cDNA are considered, PC1 and PC2 explain 37.1% and 32.4% of the variation, respectively, and breed distribution do not match grouping criteria.

Signatures of selection

Departures from neutral theory expectations were analyzed on each of the four defined breed groups ("Dairy", "Beef", "Less selected" and "Zebu") (Table 4). In the "Dairy" cattle group, these tests were

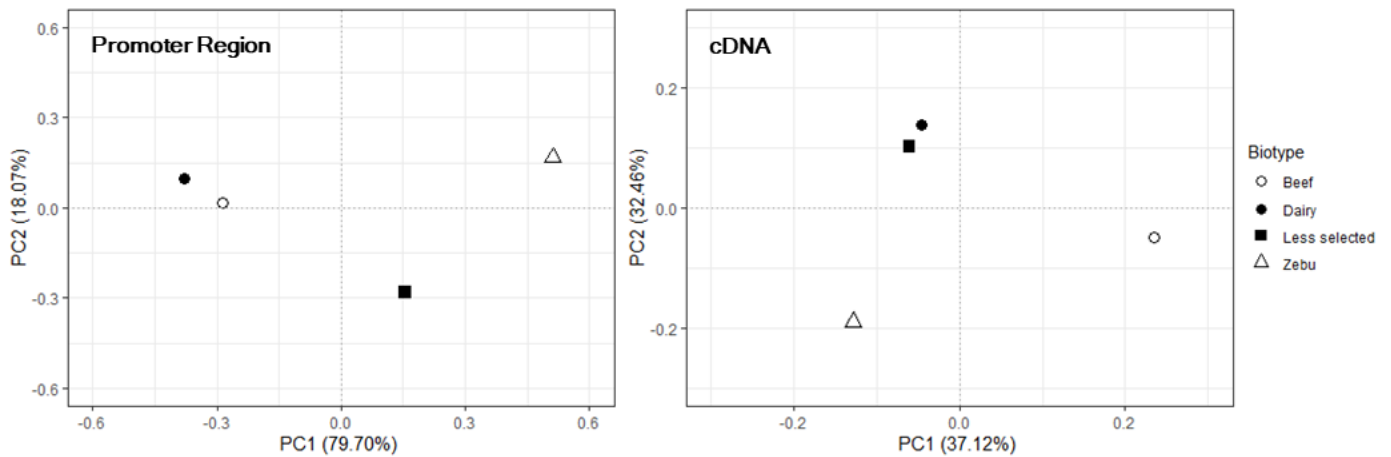


Figure 2. Bidimensional plot illustrating the results of the Principal Component Analysis performed on haplotype frequencies of the putative proximal promoter (left) and the cDNA (NM_174139.3) (right) of the bovine *PDYN* gene. The percentages of total variability explained by each PC are included between brackets. Dairy: Highly selected dairy breeds (*Bos taurus*); Beef: Highly selected beef breeds (*Bos taurus*); Less selected: Less selected breeds (*Bos taurus*), included the Auroch (*Bos primigenius*); Zebu: Zebu breeds (*Bos indicus*).

consistently negative and resulted to be statistically significant for the Genomic sequence as well as for the putative promoter region (Table 2). These results suggest that this gene and particularly the promoter region could be under positive selection. Moreover, these results pointed to a potential effect on regulatory features of *PDYN* gene expression in Dairy cattle. The other groups, on the contrary, had much higher nucleotide and haplotype diversity in the promoter and CDS than the Dairy breeds, and none of their neutrality tests were statistically significant (Table 2).

Haplotype description

Table 3 shows the mutations identified in the promoter region of the *PDYN* gene that had evidence of positive selection according to the neutrality tests, and in the CDS. These mutations define the haplotypes reported in Table 4.

The combination of 14 SNPs defined the haplotypes of the promoter region (1,000 bp) (Table 4). In this region, a total of 32 haplotypes were found among the 114 chromosomes (57 samples by 2 chromosomes each) analyzed. Sixteen of them were found in at least two chromosomes and were represented by 98 chromosomes (Table 4). The most abundant haplotype (p-Hap_1, ref; n=56) was found in the Bovine Reference Genome ARS-UCD1.2 but it was absent in the “Zebu” cattle group. In the CDS (777 bp), a total of 28 haplotypes defined by 10 SNPs were found among the 114 chromosomes analyzed. Twelve of them were found in at least two chromosomes and were represented by 98 chromosomes (Table 3). In this region, the haplotype found in the Bovine Reference Genome ARS-UCD1.2 (c-Hap_1, ref; n=15) was not the most abundant and was the only one with the “A” allele

in the SNP rs42388967 (SNP 3 in the CDS, Table 4). Although “Beef” and “Dairy” groups shared their most frequent haplotype (p-Hap_9) in the promoter that was not the case for the CDS (c-Hap_9 and c-Hap_1 for “Dairy” and “Beef” groups, respectively).

The promoter of *PDYN* had five different haplotypes in dairy breeds but with little variability, given that 34 out of 40 chromosomes shared p-Hap_1. This same haplotype was present in 20 out of 33 chromosomes of selected beef breeds, but the rest of the chromosomes of this group had seven different haplotypes in low frequencies each (Table 4). Moreover, in the group of dairy breeds, 40 out of 44 haplotypes of the CDS share the same six alleles in their 5’ end. This same trend was detected in the promoter, where 37 out of 40 haplotypes share the same seven alleles in their 3’ end. This pattern is not seen in any other breed group (Table 4). Zebu breeds had four haplotypes with a predominance of p-Hap_11 in the promoter region and showed little coincidence in haplotype distribution with the other breed groups (Table 4). Interestingly, differences between breed groups were smoothed when the CDS was considered and even the “Zebu” group showed more coincidence with the other *Bos taurus* groups. Nevertheless, for the CDS the “Dairy” group showed again the lowest variability (30 out of 44 chromosomes had Hap_9; Table 4).

The “Median Joining” network constructed using the haplotypes identified in the promoter region suggested the existence of two clades separated by three mutational steps (Figure 3A). In this network, the haplotypes found in the “Dairy” cattle group could be differentiated from those of the “Zebu” cattle group. Noteworthy, the “Dairy” group exhibited few haplotypes and they formed a star-like configuration with the reference haplotype (p-Hap_1) as central haplotype in the clade

Table 2. Diversity measures and neutrality tests. Results corresponding to the whole genomic sequence of *PDYN* and each of the different functional regions of transcript NM_174139.3 are presented for each group of breeds.

| Region | Group | n | h | Hd | S | π | θ | Tajima's D | Fu and Li's D* | Fu and Li's F* |
|---------------------------------------|---------------|-----------|-----------|----------------------|------------|----------------|----------------|--------------------|--------------------|--------------------|
| <i>PDYN</i> | | | | | | | | | | |
| Genomic sequence | Dairy | 48 | 45 | 0.997 ± 0.004 | 188 | 0.00107 | 0.00216 | -1.83842 * | -4.69877 ** | -4.33775 ** |
| | Beef | 34 | 31 | 0.989 ± 0.012 | 142 | 0.00172 | 0.00177 | -0.12272 | 0.48822 | 0.32942 |
| | Less selected | 16 | 16 | 1.000 ± 0.022 | 225 | 0.00282 | 0.00346 | -0.81225 | -0.90965 | -1.02013 |
| | Zebu | 16 | 16 | 1.000 ± 0.022 | 270 | 0.00381 | 0.00381 | -0.36633 | -0.14531 | -0.24128 |
| <i>PDYN mRNA (NM_174139.3)</i> | | | | | | | | | | |
| <i>Promoter (1,000bp)</i> | Dairy | 48 | 11 | 0.469 ± 0.090 | 13 | 0.00070 | 0.00293 | -2.30211 ** | -4.04418 ** | -4.09018 ** |
| | Beef | 34 | 7 | 0.608 ± 0.092 | 6 | 0.00143 | 0.00147 | -0.08484 | 1.20449 | 0.94936 |
| | Less selected | 16 | 9 | 0.900 ± 0.050 | 9 | 0.00249 | 0.00272 | -0.31003 | -1.01097 | -0.94107 |
| | Zebu | 16 | 7 | 0.750 ± 0.107 | 10 | 0.00158 | 0.00302 | -1.78041 # | -1.66704 | -1.95299 |
| <i>5'UTR</i> | Dairy | 48 | 2 | 0.042 ± 0.039 | 1 | 0.00024 | 0.00129 | -1.10686 | -1.82907 | -1.87498 |
| | Beef | 34 | 2 | 0.337 ± 0.083 | 1 | 0.00194 | 0.00141 | 0.56557 | 0.58040 | 0.66397 |
| | Less selected | 16 | 2 | 0.500 ± 0.074 | 1 | 0.00287 | 0.00173 | 1.30896 | 0.68829 | 0.96505 |
| | Zebu | 16 | 2 | 0.125 ± 0.106 | 1 | 0.00072 | 0.00173 | -1.16221 | -1.45287 | -1.56820 |
| <i>CDS</i> | Dairy | 48 | 10 | 0.598 ± 0.079 | 9 | 0.00157 | 0.00261 | -1.12637 | -1.91413 | -1.95186 # |
| | Beef | 34 | 15 | 0.836 ± 0.056 | 8 | 0.00296 | 0.00252 | 0.52289 | 1.31840 | 1.25513 |
| | Less selected | 16 | 11 | 0.950 ± 0.036 | 9 | 0.00326 | 0.00349 | -0.24232 | 0.42855 | 0.28108 |
| | Zebu | 16 | 10 | 0.942 ± 0.036 | 7 | 0.00347 | 0.00272 | 0.98621 | 1.31791 | 1.40942 |
| <i>3'UTR</i> | Dairy | 48 | 26 | 0.926 ± 0.028 | 35 | 0.00472 | 0.00497 | -0.17413 | -1.84513 | -1.48846 |
| | Beef | 34 | 16 | 0.775 ± 0.074 | 18 | 0.00292 | 0.00278 | 0.16929 | 0.90055 | 0.78173 |
| | Less selected | 16 | 15 | 0.992 ± 0.025 | 22 | 0.00539 | 0.00477 | 0.28355 | 0.46251 | 0.75674 |
| | Zebu | 16 | 14 | 0.975 ± 0.035 | 18 | 0.00491 | 0.00342 | 1.73346 | 1.01041 | 1.39731 |

n: number of chromosomes; h: number of haplotypes; Hd: haplotype diversity; S: number of segregating sites; π : nucleotide diversity (per site); $\theta = 4N\mu$ (N and μ are the effective population size, and the mutation rate per DNA sequence per generation, respectively); Dairy: Highly selected dairy breeds (*Bos taurus*); Beef: Highly selected beef breeds (*Bos taurus*); Less selected: less selected breeds (*Bos taurus*), includes the Auroch (*Bos primigenius*); Zebu: zebu breeds (*Bos indicus*). #0.1>p>0.05; *p<0.05; **p<0.01. A region of the gene was considered under selection when both Tajima's and Fu & Li's neutrality tests produced significant results. These regions are shown in bold. 5'UTR and 3'UTR: untranslated regions; CDS: coding sequence.

(Figure 3A). In contrast, the haplotypes from the other groups were sparsely distributed. On the other hand, the “Median Joining” network constructed using the haplotypes identified in the CDS did not exhibit a clear pattern of variation and contrary to what happened in the promoter region, the haplotypes found in the CDS in the different cattle groups were not separated in this network (Figure 3B).

The 28 haplotypes identified in the CDS (Table 4) could be translated into six propeptide sequences, which had

no more than four changes compared to the translated reference sequence. The four breed groups coincidentally had the same translated sequence (translated sequence of c-Hap_9) as the most abundant (Table 4). Interestingly, none of the aminoacid substitutions affected the sequence of the functional peptides that are processed from the translated propeptide (Table 3, Figure 1).

Table 3. List of mutations identified in two regions of the *PDYN* gene (promoter and coding sequence, CDS).

| Region | Position | ID | Mutation | aa position | aa substitution | Coordinates (ARS-UCD 1.2) |
|---|----------|--------------|----------|-------------|-----------------|---------------------------|
| <i>PDYN</i> mRNA promoter (1,000 bp) | 1 | rs483143590 | C>T | | | 53,133,835 |
| | 2 | rs133702134 | C>T | | | 53,133,850 |
| | 3 | | T>C | | | 53,133,873 |
| | 4 | | ->T | | | 53,133,905 |
| | 5 | | G>A | | | 53,133,933 |
| | 6 | | C>G | | | 53,133,938 |
| | 7 | | G>T | | | 53,133,945 |
| | 8 | | T>- | | | 53,134,299 |
| | 9 | | C>- | | | 53,134,300 |
| | 10 | | C>T | | | 53,134,467 |
| | 11 | rs480238520 | C>A | | | 53,134,474 |
| | 12 | rs445119268 | ->C | | | 53,134,494 |
| | 13 | | A>C | | | 53,134,596 |
| | 14 | | T>G | | | 53,134,604 |
| CDS (777 bp) | 1 | rs518871097 | C>T | 15 | | 53,146,628 |
| | 2 | rs211042415 | T>C | 52 | | 53,146,739 |
| | 3 | rs42388967 | A>G | 75 | R>G | 53,146,806 |
| | 4 | rs1116570653 | C>A | 76 | L>I | 53,146,809 |
| | 5 | rs473280776 | A>G | 114 | N>S | 53,146,924 |
| | 6 | rs718114917 | C>T | 127 | A>V | 53,146,963 |
| | 7 | rs132886325 | G>A | 127 | | 53,146,964 |
| | 8 | rs524857811 | C>T | 154 | | 53,147,045 |
| | 9 | rs134167025 | G>A | 160 | A>T | 53,147,061 |
| | 10 | rs42388969 | A>C | 177 | | 53,147,114 |

The "Position" column indicates the position in the corresponding haplotypes described in Table 4.

DISCUSSION

The involvement of Endogenous Opioid Peptides (EOP) as negative regulators of the reproductive cycle has been known for a long time, but their role has not been completely elucidated. In cattle, EOP received attention as factors consistently involved in the inhibitory effects of suckling on reproductive activity (Malven *et al.*, 1986). Frequent suckling or milking increases levels of EOP in the brain, which in turn increases the sensitivity of the hypothalamus to the negative feedback from estrogen (Squires, 2010). This experimental evidence confirms the role of Dynorphin A and EOP in general, as inhibitors of reproductive activity in mammals, and justify the search of selection signatures conducted in the present work to explain the well-known variability in reproductive

performance among cattle breeds, that is noticeable since the onset of puberty (Diskin and Kenny, 2014).

In the present experiment, we analyzed an annotated sequence of *PDYN*, which encoded, among others, the endogenous opioid peptide Dynorphin A (Figure 1), from the last version of the Bovine Reference Genome (Genbank accession NM_174139.3). This sequence has a genomic organization that is consistent with the cDNA originally cloned by Jiang *et al.* (1997); Genbank accession U58500.1).

The dataset analyzed here included the only available sample from the Auroch (*Bos primigenius*, Park *et al.*, 2015) that were grouped with less selected breeds. This sample did not show marked differences with other groups. However, it must be noted that a single sample is not enough to identify signatures of selection. Thus,

Table 4. List of haplotypes. List of haplotypes defined by mutations identified in the coding sequence (CDS) and in the promoter region of the *PDYN*. Only haplotypes found in at least two chromosomes are shown.

| Region | Haplotype | n | D | B | L_S | Z | n changes | Description |
|---|-----------|----|----|----|-----|----|-----------|----------------------------------|
| PDYN mRNA promoter (1,000bp) | p-Hap_1 | 56 | 34 | 20 | 2 | | reference | CCT- ACTT CCC-AT |
| | p-Hap_11 | 9 | | | 1 | 8 | 4 | CTT-GCT --CC-AT |
| | p-Hap_7 | 5 | | 2 | 3 | | 2 | CTT-GCT TCCC-AT |
| | p-Hap_8 | 4 | 1 | 3 | | | 1 | CCT- ACG TCCC-AT |
| | p-Hap_2 | 2 | | 2 | | | 7 | CTTTGCT --CA-AG |
| | p-Hap_3 | 2 | | 1 | 1 | | 3 | CCT- ACG --CC-AT |
| | p-Hap_4 | 2 | | 1 | 1 | | 2 | CCT- ACT --CC-AT |
| | p-Hap_5 | 2 | | 2 | | | 3 | CTT-GCT TCCA-AT |
| | p-Hap_9 | 2 | | 2 | | | 2 | CCT- ACG TCTC-AT |
| | p-Hap_12 | 2 | | | 1 | 1 | 4 | CCT- ACG -- TC -AT |
| | p-Hap_16 | 2 | 1 | | 1 | | 1 | CCT- ACTT TCTC-AT |
| | p-Hap_17 | 2 | | | 2 | | 2 | CCT- ACG TCTCCAT |
| | p-Hap_18 | 2 | 2 | | | | 1 | CCT- ACTT CCC- CT |
| | p-Hap_23 | 2 | 2 | | | | 1 | TCT - ACTT CCC-AT |
| | p-Hap_30 | 2 | | | | | 2 | CTC-GCT --CC-AT |
| | p-Hap_31 | 2 | | | | | 2 | CTT-GGT --CC-AT |
| Totals | 98 | 40 | 33 | 12 | 13 | | | |
| CDS (777 bp) | c-Hap_1 | 15 | 1 | 13 | 1 | | reference | CTACACGCGA |
| | c-Hap_9 | 36 | 30 | 2 | 2 | 2 | 3 | CT GCG CGCGC |
| | c-Hap_2 | 11 | 3 | 5 | | 3 | 4 | CCGCG CGCGC |
| | c-Hap_3 | 10 | 4 | 3 | 3 | | 2 | CT GCG CGCGA |
| | c-Hap_12 | 9 | 5 | 1 | 1 | 2 | 4 | CT GCG CACAA |
| | cHap_15 | 4 | 1 | 1 | 2 | | 3 | CT GCG CACGA |
| | c-Hap_16 | 3 | | | 1 | 2 | 3 | CT GCG CG CAA |
| | c-Hap_10 | 2 | | 1 | 1 | | 1 | CT GCA CGCGA |
| | c-Hap_13 | 2 | | 1 | | | 1 | CCGCG CGCGA |
| | c-Hap_17 | 2 | | | 1 | 1 | 6 | TTGCGT GCAC |
| | c-Hap_18 | 2 | | | 2 | | 5 | CT GAG CACAA |
| | c-Hap_28 | 2 | | | | 2 | 5 | CCGCG CG TGC |
| | Totals | 98 | 44 | 27 | 14 | 13 | | |

The total number of identified chromosomes (n) are discriminated by breed group. Dairy: Highly selected dairy breeds (*Bos taurus*); Beef: Highly selected beef breeds (*Bos taurus*); Less selected: less selected breeds (*Bos taurus*), includes the Auroch (*Bos primigenius*); Zebu: zebu breeds (*Bos indicus*). n changes: number of changes respect to the bovine reference genome ARS-UCD1.2. Differences respect to the reference genome are shown in bold. The positions of SNPs that generate aa substitutions in the CDS are in bold italic font.

more genomes from this group should be sequenced to be more conclusive about the effects of domestication (Orlando, 2015).

The PCA results presented in Figure 2, showing sample substructure differed depending in the analyzed regions of *PDYN* (promoter and cDNA) indicates that the observed differences are not only justified by the well-known genetic distance between *Bos indicus* and *Bos taurus* breeds, product of sub-speciation, and justify the definition of four separated groups used in the other analyses. The comparison of haplotype frequencies among breed groups for both genomic regions (Table 4, Figure 3) also supports the existence of directional selection on regulatory regions of the gene and not only random genetic drift associated with the breed or subspecies formation in the definition of sequence variation.

It could be argued that low genetic variability is the result of reduced effective population size (N_e), a feature common to selected populations in most livestock species (Taylor *et al.*, 2016). In this case, the group of dairy breeds was integrated by two different breeds, Holstein and Jersey that share many selection objectives and also a common production system. Also, this group had the largest sample size.

The methods that were used in the present work are appropriate to identify recent selection (Biswas and Akey, 2006). Indeed, the results of the Tajima's D (Tajima, 1989) and Fu & Li D^* and F^* (Fu and Li, 1993) exhibited statistically significant negative values in the "Dairy" group (Table 2) which pointed out to a recent selection process in modern, highly specialized breeds. This evidence of recent selection is consistent with the

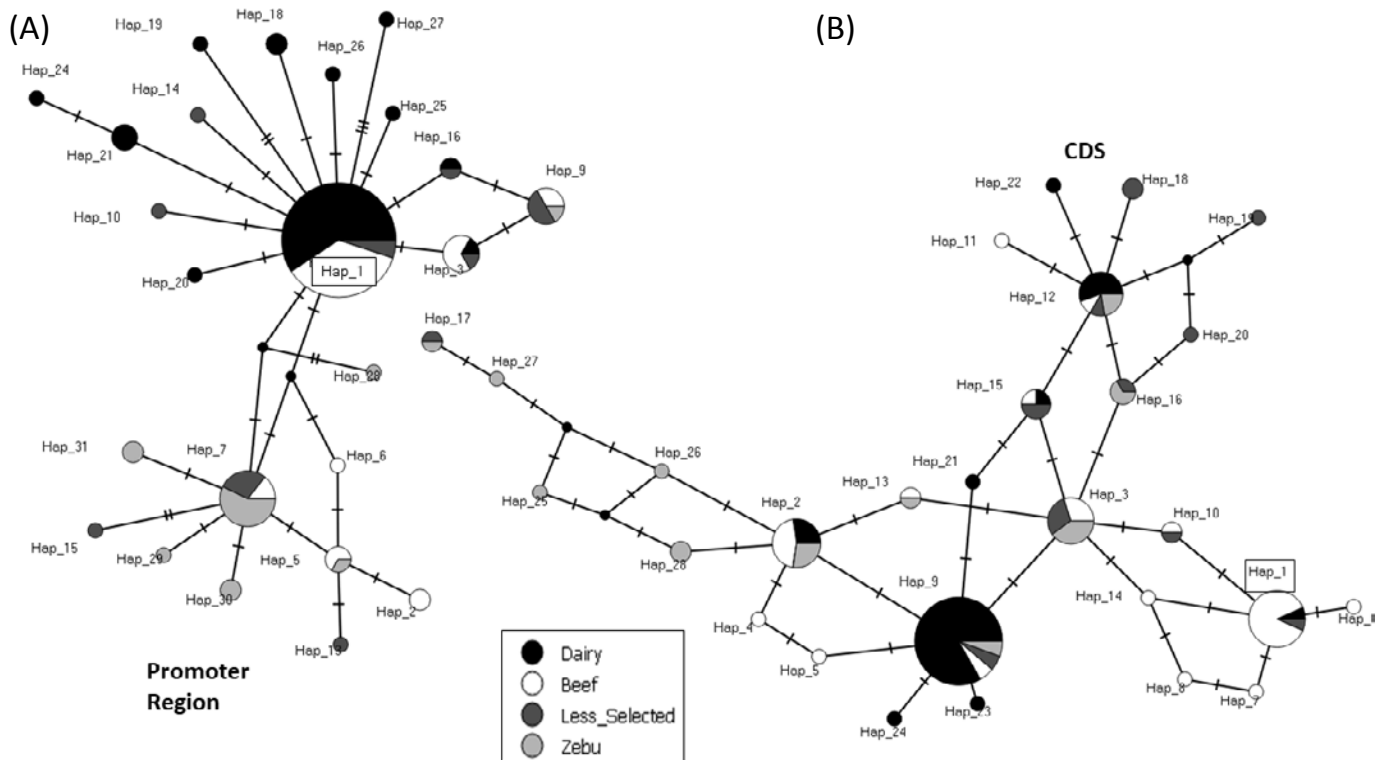


Figure 3. Median Joining network constructed with the haplotypes corresponding to (A) the putative proximal promoter (1,000 bp) and (B) the coding sequence (CDS) of *PDYN* gene. The size of each circle is proportional to the corresponding chromosome frequency. Slashes represent the number of mutations separating each haplotype. In each network Hap_1 (box) corresponds to the Bovine Reference Genome ARS-UCD1.2. Dairy: Highly selected dairy breeds (*Bos taurus*); Beef: Highly selected beef breeds (*Bos taurus*); Less selected: Less selected breeds (*Bos taurus*), included the Auroch (*Bos primigenius*); Zebu: Zebu breeds (*Bos indicus*).

emphasis of intense artificial selection on production traits, particularly those related to reproductive efficiency. A compelling argument in favor of positive selection on a gene such as *PDYN* comes from the intrinsic characteristics of different livestock production systems. Under extensive conditions, the cow not only nurses but also protects the progeny, and the maternal bond exerts a strong inhibiting effect on reproductive activity (Williams, 1990). In more intensified systems such as those for dairy production the calf is separated from the mother hours after parturition. Moreover, dairy cows are usually expected to get pregnant under a strongly negative energy balance (Butler and Smith, 1989; Beam and Butler, 1999; Vercouteren *et al.*, 2015). Clearly, in these extreme cases, the physiological and environmental cues that have to be processed at the hypothalamic level to regulate reproductive cycles are very different. It could be hypothesized that in females from more intensified production systems, the effects of some inhibitory mechanisms acting on the regulation of reproduction are relaxed, favoring the selection response for reproductive efficiency. The results reported here showed that the “Dairy” group presented fewer haplotypes than the other cattle groups

in the promoter region and that these haplotypes exhibit low differences among them (Table 4, Figure 3A). A detailed description on the epigenetic regulation of the expression on *KISS1* and *TAC3* by implementing histone modifications was recently made by Toro *et al.* (2018). However, it does not seem to be the case of *PDYN* gene, the third member of the triad of the “KNDy hypothesis” (Smith *et al.*, 2014). The results presented here warrants further investigation on the mechanisms underlying the *PDYN* transcriptional regulation.

The results presented here suggest that there is selection pressure acting on putative regulatory regions on the proximal promoter of *PDYN* (Table 2), which seems to be a common feature of selection for quantitative or complex traits. Modifications of the coding sequence are more common in the case of genes underlying mendelian traits, in which the mutation has a strong effect on the phenotype (Boyle *et al.*, 2017). Although aminoacid substitutions on the propeptide were detected in the present analysis, they do not seem to affect the sequence of functional peptides, stressing their biological importance (Table 3, Figure 1). Also, a not clear pattern of differentiation was observed among the haplotypes of the four cattle groups in the coding

sequence (Figure 3B). Selection on regulatory regions of *PDYN* would affect the expression of all the different peptides coded by the gene. Given the physiological role of Dynorphin A, the interest to improve reproductive efficiency in highly specialized cattle breeds provides a very reasonable explanation to justify selection on *PDYN*. Nevertheless, at this point, the relevance of the other peptides cannot be ruled out. Further research should clarify the role of EOP, particularly Dynorphin A, in the regulation of reproduction both within and between cattle breeds.

The usefulness of QTN identification (Quantitative Trait Nucleotides underlying Quantitative Trait Loci) in animal breeding is a matter of debate. Present genomic selection in cattle is mostly based on dense arrays of anonymous markers; however, methods are being developed to include QTN information to improve prediction accuracy (Fragomeni *et al.*, 2017). Besides, there are initiatives to enhance progress in animal selection through the combination of conventional breeding and gene editing (*e.g.*: Promotion of Alleles by Genome Editing; Jenko *et al.*, 2015). However, there are doubts about the efficacy of this approach due to the intrinsic polygenic basis of quantitative variation (Simianer, 2018). Independently of potential direct applications of the knowledge about *PDYN* variation among cattle breeds, the present work contributed with elements for a deeper understanding of the complex interaction among genetic variation, artificial selection, and environmental effects.

BIBLIOGRAPHY

- Amstalden M., Williams G.L. (2015) Neuroendocrine Control of Estrus and Ovulation. In: Hopper R.M. (Ed.) Bovine Reproduction. Wiley Blackwell, Starkville, Mississippi, USA, pp. 203-219.
- Atkins J.A., Pohler K.G., Smith M.F. (2013) Physiology and Endocrinology of Puberty in Heifers. *Vet. Clin. North. Am. Food. Anim. Pract.* 29: 479-492.
- Bandelt H.J., Forster P., Röhl A. (1999) Median-Joining Networks for Inferring Intraspecific Phylogenies. *Mol. Biol. Evol.* 16: 37-48.
- Beam S.W., Butler W.R. (1999) Effects of energy balance on follicular development and first ovulation in postpartum dairy cows. *J. Reprod. Fertil. Suppl.* 54: 411-424.
- Biswas S., Akey J.M. (2006) Genomic insights into positive selection. *Trends Genet.* 22: 437-446.
- Boyle E.A., Li Y.I., Pritchard J.K. (2017) An expanded view of complex traits: from polygenic to omnigenic. *Cell.* 169: 1177-1186.
- Butler W.R., Smith R.D. (1989) Interrelationships Between Energy Balance and Postpartum Reproductive Function in Dairy Cattle. *J. Dairy Sci.* 72: 767-783.
- Corva P.M., Villarreal E.L., Mezzadra C.A. (1995) Reproductive traits of Angus, Criollo and reciprocal crossbred females in the temperate area of Argentina. *Anim. Sci.* 61: 241-249.
- Daetwyler H., Capitan A., Pausch H. (2014) Whole-genome sequencing of 234 bulls facilitates mapping of monogenic and complex traits in cattle. *Nat. Genet.* 46: 858-865.
- Day R., Lazure C., Basak A. (1998) Prodynorphin Processing by Proprotein Convertase 2. *J. Biol. Chem.* 273: 829-836.
- Diskin M.G., Kenny D.A. (2014) Optimising reproductive performance of beef cows and replacement heifers. *Animal* 8: 27-39.
- Ferraz Jr. M.V.C., Pires A.V., Santos M.H. (2018) A combination of nutrition and genetics is able to reduce age at puberty in Nelore heifers to below 18 months. *Animal* 12: 569-574.
- Fragomeni B.O., Lourenco D.A.L., Masuda Y. (2017) Incorporation of causative quantitative trait nucleotides in single-step GBLUP. *Genet. Sel. Evol.* 49: 59.
- Freetly H.C., Kuehn L.A., Cundiff L.V. (2011) Growth curves of crossbred cows sired by Hereford, Angus, Belgian Blue, Brahman, Boran, and Tuli bulls, and the fraction of mature body weight and height at puberty. *J. Anim. Sci.* 89: 2373-2379.
- Fu Y.X., Li W.H. (1993) Statistical tests of neutrality of mutations. *Genetics* 133: 693-709.
- Goodman R.L., Lehman M.N., Smith J.T. (2007) Kisspeptin neurons in the arcuate nucleus of the ewe express both dynorphin A and neurokinin B. *Endocrinology* 148: 5752-5760.
- Heaton M.P., Smith T.P.L., Carnahan J.K. (2016) Using diverse U.S. beef cattle genomes to identify missense mutations in EPAS1, a gene associated with high-altitude pulmonary hypertension. *F1000Research* 5: 2003.
- Irano N., De Camargo G.M.F., Costa R.B. (2016) Genome-wide association study for indicator traits of sexual precocity in Nelore cattle. *PLoS One* 11: 1-14.
- Jenko J., Gorjanc G., Cleveland M.A. (2015) Potential of promotion of alleles by genome editing to improve quantitative traits in livestock breeding programs. *Genet. Sel. Evol.* 47: 55.
- Jiang H., Weesner G.D., Malven P.V. (1997) cDNA sequence and expression of bovine prodynorphin. *Gene* 186: 279-283.
- Kimura M. (1968) Evolutionary Rate at the Molecular Level. *Nature* 217: 624-626.
- Larsson A. (2014) AliView: a fast and lightweight alignment viewer and editor for large datasets. *Bioinformatics* 30: 3276-3278.
- Laster D.B., Smith G.M., Cundiff L.V. (1979) Characterization of biological types of cattle (Cycle II). II Postweaning growth and puberty of heifers. *J. Anim. Sci.* 48: 500-508.
- Lehman M.N., Coolen L.M., Goodman R.L. (2010) Minireview: Kisspeptin/neurokinin B/dynorphin (KNDy) cells of the arcuate nucleus: A central node in the control of gonadotropin-releasing hormone secretion. *Endocrinology* 151: 3479-3489.
- Leigh J., Bryant D. (2015) PopART: Full-feature software for haplotype network construction. *Methods Ecol. Evol.* 6: 1110-1116. Available at: <http://popart.otago.ac.nz>.
- Li H. (2011) A statistical framework for SNP calling, mutation discovery, association mapping and population genetical parameter estimation from sequencing data. *Bioinformatics* 27: 2987-2993.
- Malven P.V. (1986) Inhibition of pituitary LH release resulting from endogenous opioid peptides. *Domest. Anim. Endocrinol.* 3: 135-144.
- Malven P.V., Parfet J.R., Gregg D.W. (1986) Relationships among Concentrations of Four Opioid Neuropeptides and Luteinizing

- Hormone-Releasing Hormone in Neural Tissues of Beef Cows Following Early Weaning. *J. Anim. Sci.* 62: 723-733.
- Meirelles C.F., Abdalla A.L., Vitti D.M.S.S. (1994) The effect of feed supplementation on the onset of puberty in Brazilian dairy heifers. *Sci. Agric.* 51: 374-380.
- Merchant S., Wood D.E., Salzberg S.L. (2014) Unexpected cross-species contamination in genome sequencing projects. *Peer J.* 2: e675.
- Orlando L. (2015) The first aurochs genome reveals the breeding history of British and European cattle. *Genome Biol.* 16: 225.
- Pardo A.M., Villarreal E.L., Papaleo Mazzucco J. (2018) Sexual precocity and productivity of beef cattle female under grazing conditions. *Anim. Prod. Sci.* 59: 757-766.
- Park S.D.E., Magee D.A., McGettigan P.A. (2015) Genome sequencing of the extinct Eurasian wild aurochs, *Bos primigenius*, illuminates the phylogeography and evolution of cattle. *Genome Biol.* 16: 234.
- Pedersen B.S., Quinlan A.R. (2018) Mosdepth: quick coverage calculation for genomes and exomes. *Bioinformatics* 34: 867-868.
- R Development Core Team (2018) R: a language and environment for statistical computing Vienna, Austria: The R Foundation for Statistical Computing. Available at: <http://www.r-project.org/>.
- Rozas J., Ferrer-Mata A., Sanchez DelBarrio J.C. (2017) DnaSP 6: DNA sequence polymorphism analysis of large data sets. *Mol. Biol. Evol.* 34: 3299-3302.
- Sartori R., Gimenes L.U., Monteiro Jr. P.L.J. (2016) Metabolic and endocrine differences between *Bos taurus* and *Bos indicus* females that impact the interaction of nutrition with reproduction. *Theriogenology* 86: 32-40.
- Scheet P., Stephens M. (2006) A Fast and Flexible Statistical Model for Large-Scale Population Genotype Data: Applications to Inferring Missing Genotypes and Haplotypic Phase. *Am. J. Hum. Genet.* 78: 629-644.
- Simianer H. (2018) Of Cows and Cars. *J. Anim. Breed. Genet.* 135: 249-250.
- Smith M.F., Hawken P.A.R., Lehman M.N. (2014) The role of kisspeptin in reproductive function in the ewe. In: Juengel J.L., Miyamoto A., Price C., Reynolds L.P., Smith M.F., Web R. (Eds.) *Reproduction in Domestic Ruminants VIII*. Context, UK, pp. 105-116.
- Squires E.J. (2010) *Applied Animal Endocrinology*. 2nd Ed. CABI Publishing, Guelph, Ontario, Canada.
- Tajima F. (1989) Statistical Method for Testing the Neutral Mutation. *Genetics* 123: 585-595.
- Taylor J.F., Taylor K.H., Decker J.E. (2016) Holsteins are the genomic selection poster cows. *Proc. Natl. Acad. Sci.* 113: 7690-7692.
- Toro C.A., Wright H., Aylwin C.F. (2018) Trithorax dependent changes in chromatin landscape at enhancer and promoter regions drive female puberty. *Nat. Commun.* 9: 57.
- Vercouteren M.M.A.A., Bittar J.H.J., Pinedo P.J. (2015) Factors associated with early cyclicity in postpartum dairy cows. *J. Dairy Sci.* 98: 229-239.
- Weems P.W., Lehman M.N., Coolen L.M. (2018) The Roles of Neurokinins and Endogenous Opioid Peptides in Control of Pulsatile LH Secretion. In: Litwack G. (Ed.) *Vitamins and Hormones*. Elsevier Inc., Academic Press, pp. 89-135.
- Wickham H. (2016) *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag, New York.
- Williams G.L. (1990) Suckling as a regulator of postpartum rebreeding in cattle: a review. *J. Anim. Sci.* 68: 831-852.
- Zimin A.V., Delcher A.L., Florea L. (2009) A whole-genome assembly of the domestic cow, *Bos taurus*. *Genome Biol.* 10: R42.